STRUCTURE OF SIBIRINONE, A NEW α-PYRONE FROM HYPOMYCES*

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A new α-pyrone, sibirinone, was isolated from a culture of Hypomyces semitranslucens G. Arnold, the imperfect state of which is Sibirina fungicola G. Arnold. Sibirinone, $C_8H_8O_2$, MW 136.0531, has λ_{mux} 333 (4.15) and 226 nm (4.25) and v_{max} at 1740, 1715 and 1653, 1605, 1550 cm⁻¹. The UV and IR spectra were very similar to those of other extended α -pyrones, [1, 2] suggesting structure 1 for sibirinone. The NMR spectrum showed signals at 1.87 (3H, dd, J = 7.0, 1.2) for the methyl protons, 5.97 (1H, d, J = 6.5) for the C₅ protons, 6.03 (1H, dq, J = 15, 1.2) for the C_7 proton, 6.12 (1H, d, J = 9.5) for the C_3 proton, 6.67 (1H, dq, J = 15, 7) for the C₈ proton, and 7.28 (1H, dd, J = 6.5, 9.5) for the C_4 proton. Signals for the C_3 , C₇ and C₈ protons were crowded together and the doublet of the quartet for the C₇ proton was partially obscured. However, on adding Eu (DPM)3, as expected, the signals for the C₃, and to a lesser extent that for the C₅ proton, shifted down field and the C_7 proton signals, which did not shift, were clearly visible. The coupling pattern and constants of the side chain are comparable to those of the trans-2-pentenyl A₃XY system of trans-crotonic acid and similar compounds [3]. The ring proton shifts show that the substitution is on carbon six and the coupling constants this should be corrected to read $J_{3,4} = 9.5$; $J_{4,5} = 6.5 (J_{3,4} = 6.5, J_{4,5} = 9.5)$ are characteristic of α-pyrones [4].

The mass spectral fragmentation pattern of sibirinone was in complete agreement with this structure: [5]: peaks at 136 (M, 50%), 108 (M—CO, 95%), 95 (M—C₃H₃. 40%) and 89 (M—CO—CHO, 100%). Sibirinone was hydrogenated in ethanol over platinum catalyst. The main product separated by preparative TLC, had MS peaks at 144 (M), 99, 73 and 60 (base peak). This compound was identified as *n*-octanoic acid by comparison of its IR spectrum with that of an authentic sample. Formation of the octanoic acid on hydrogenation,

further confirms the structure of sibirinone as 1. Sibirinone was not active against the strain of Staphylococcus aureus (ATCC 9144) used in our serial dilution tests.

EXPERIMENTAL

General procedures. Mps are uncorr. PMR spectra were taken in CDCl₃ with TMS as internal standard and are expressed in the δ scale.

Isolation of stbirinone. H. semitranslucens was grown in still culture in a dextrose-yeast medium in Fernbach flasks in the dark at 25°. The culture was harvested after four weeks. The culture liquid was extracted with EtOAc and the extract was taken to dryness in vacuo. The residue was then chromatographed on a Si gel column (50 times the weight) and sibirinone was eluted with petrol-EtOAc (1:1) and was crystallised from Et₂O-petrol (1:1), mp 58-59°.

Hydrogenation of sibirinone. Sibirinone (10 mg) was hydrogenated in the presence of reduced PtO (15 mg) in EtOH. The products were separated by preparative TLC. The main component was n-octanoic acid.

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REFERENCES

- Nair, M. S. R. and Carey, S. T. (1975) Tetrahedron Letters 19, 1655.
- Scott, A. I. (1964) Interpretation of Ultraviolet Spectra of Natural Products p, 141. Pergamon Press, New York.
- Kowalewski, V. J. and De Kowalewski, D. G. (1960) J. Chem. Phys. 33, 1794.
- Batterham, T. J. (1973) NMR Spectra of Simple Heterocycles, p. 392. John Wiley, New York.
- Budzikiewicz, H., Djerassi, C. and Williams, D. H. (1967)
 Mass Spectrometry of Organic Compounds, pp. 208-210.
 Holden-Day, San Francisco.

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